



Thermo Scientific **Synchronis HPLC Columns**



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When developing a new method, one of the most important goals for the chromatographer is to achieve a consistent, reproducible separation. The selection of a highly reproducible HPLC column is essential if this goal is to be attained.

Our Thermo Scientific™ Synchronis™ HPLC columns are manufactured, packed and tested in ISO9000:2008 accredited facilities. Each lot of silica is tested for the physical properties of the silica support and only released for production if it meets the stringent test specifications.

Each bonded lot of chromatographic packing material is rigorously tested for primary and secondary interactions with the bonded phase.

New, enhanced, automated packing methods drive consistency even further and every column is individually tested to ensure that it meets the required quality.

These extensive testing and quality control procedures ensure the delivery of a consistent product, column after column.

**Predictability you can count on,
it's a beautiful thing.**



Synchronis HPLC columns are available in a range of chemistries to give reproducible separations in reversed phase, HILIC and normal phase chromatography.

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Synchronis Stationary Phases

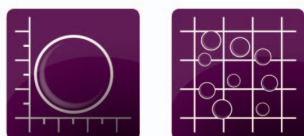
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Testing and Quality Control

Silica Characterization

Consistent separations require a rugged, reproducible silica backbone and to achieve this consistency, strict control of the physical properties of the silica particles is essential. Each manufactured lot of Synchronis silica is tightly controlled and extensively tested for its particle size and distribution, pore size and surface area and elemental purity. Only those batches which meet the rigorous quality control specifications are used.

Particle size and distribution



Tight control of the particle classification process ensures that a narrow particle size distribution is achieved around the target particle size, an important consideration for consistent chromatographic efficiency. Each manufactured lot of Synchronis silica is tested for its particle size and distribution using a laser particle size analyser. Three particle sizes are available: 1.7 μm for rapid UHPLC separations plus 3 μm and 5 μm for the more traditional HPLC analysis.

Pore size and surface area



Well controlled pore size and surface area are key to ensuring consistent carbon load and retentive properties of the chromatographic media. Each batch of Synchronis silica is tested for its pore size and surface area using liquid nitrogen adsorption.

Synchronis columns are based on highly pure 100 \AA silica, with a surface area of 320 m^2/g , compared to 200 m^2/g for typical silica based material. This greater surface area ensures good retention of analytes having a range of hydrophobicity, away from the solvent front. The high surface area also allows for higher sample loading.

Silica purity (metals content)

The purity of the silica support is of particular importance when considering the separation of polar and basic compounds. Older, less pure silica supports contain a greater number of metallic impurities. The presence of certain metallic impurities with electron withdrawing properties (particularly aluminium) in silica can activate the silanols so that they become highly acidic, which can lead to peak tailing for basic solutes. Metallic impurities can also complex with chelating solutes, resulting in asymmetrical or tailing peaks. In extreme cases, these interactions may be strong enough to result in complete retention of the solute.

Each batch of Synchronis silica is tested for metals content using atomic emission spectroscopy. The specification and typical results are shown below.

Metal	Na (ppm)	Mg (ppm)	Al (ppm)	Ca (ppm)	Fe (ppm)	Zr (ppm)
Specification	<10	<10	<10	<10	<10	<10
Batch A	0.9	0.3	<1	<0.5	0.9	0.5
Batch B	0.8	<0.1	<1	<0.5	0.9	1.1
Batch C	1.1	<0.1	<1	<0.5	1.5	1.4
Batch D	1.1	<0.1	<1	<0.5	0.8	1.4
Batch E	1.3	0.4	1.4	0.8	1.1	1.2

Bonded Phase Characterization

Synchronis HPLC columns are bonded and endcapped with a range of stationary phases to effect different selectivity in separation. Whatever the bonded phase, rigorous testing and precise control of the bonding process are essential to achieve consistent chromatography. Each batch of Synchronis chromatographic media is tested for carbon load and characterized by stringent chromatographic testing before it is used to pack columns.

Carbon loading and surface coverage



The hydrophobic retention of a stationary phase is directly dependent on the carbon loading on the silica. Precise control of the batch to batch carbon loading is therefore a critical factor in ensuring consistent retention times. Synchronis reversed phase columns are densely bonded and double endcapped to minimise the number of residual silanols available to interact with basic analytes. Each batch of bonded phase is tested for carbon load using a total carbon analyzer.

Chromatographic tests

The retention properties of a reversed-phase packing material can be categorized into hydrophobic interactions, which include the measure of the hydrophobicity of the ligand and its density, steric or shape selectivity and secondary interactions such as silanol and surface metal activity. The impact that interactions between analytes and silanols have on the chromatographic performance depends on the pH of the mobile phase. Silanols on the silica surface can hydrogen bond (both as a donor and acceptor) and dissociated silanols can ion exchange with protonated bases.

To ensure consistent, predictable separations, the chromatographic media packed into Synchronis HPLC columns is extensively characterized using a series of diagnostic chromatographic tests, based on those developed by Tanaka¹. These tests rigorously probe interactions between analytes and stationary phase, measuring hydrophobicity, shape selectivity and secondary interactions with bases, acids and chelators.

1. K. Kimata, K. Iwaguchi, S. Onishi, K. Jinno, R. Eksteen, K. Hosoya, M. Araki and N. Tanaka, J.Chromatographic Science, v. 27 p. 721-728 (1989)

Testing and Quality Control

Test 1: Hydrophobic Interactions

Hydrophobic Retention

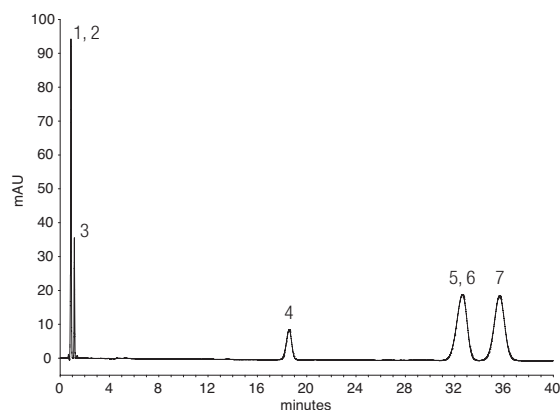


The first hydrophobic interaction that is measured is Hydrophobic Retention (HR). The capacity factor of a neutral, hydrophobic hydrocarbon (pentylbenzene) gives a measure of the hydrophobicity of the stationary phase and therefore a measure of the carbon load on the silica.

Hydrophobic Selectivity

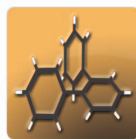


The second measurement of hydrophobic interaction that is carried out is Hydrophobic Selectivity (HS). The selectivity factor between pentylbenzene and butylbenzene provides a measure of the surface coverage of the phase; these two alkylbenzenes differ by one methylene group and their selectivity is dependent on the density of the bonded ligand.



Example chromatogram for Test 1: Hydrophobic interactions

Steric Selectivity



Steric Selectivity (SS) is the measurement of the ability of the stationary phase to distinguish between molecules with similar structures and hydrophobicity but different shapes. The selectivity factor between o-terphenyl and triphenylene is indicative of steric selectivity as the former has the ability to twist and bend, while the latter has a rigid structure and is retained quite differently.

Hydrogen Bonding Capacity

The Hydrogen Bonding Capacity (HBC) of the stationary phase is probed by determining the selectivity factor between caffeine and phenol. This provides a measure of the number of available silanol groups and the degree of endcapping. A low value indicates a low level of silanols available to hydrogen bond with caffeine and therefore an indication of the thoroughness of the endcapping.

Column: Syncronis C18 5 μm , 100 mm x 4.6 mm
Mobile phase: water:methanol (35:65)
Flow rate: 1.0 mL/min
Temperature: 40 °C
Detection: 254 nm
Injection volume: 10 μL

1. Theophylline
2. Caffeine
3. Phenol
4. Butylbenzene
5. o-Terphenyl
6. Pentylbenzene
7. Triphenylene

Test 2: Interactions with Basic and Chelating Compounds

Activity towards basic compound

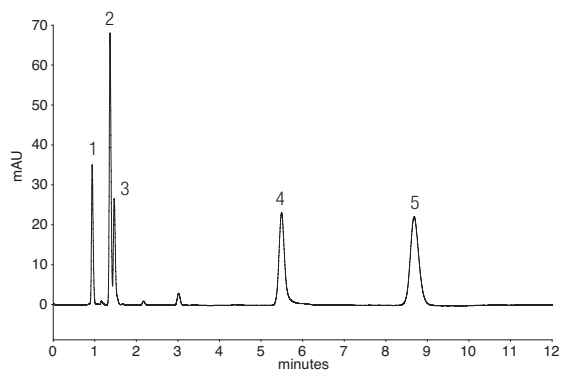
The presence of dissociated silanols at pH>7 can cause poor peak shapes of protonated basic compounds, such as amitriptyline. Secondary ion exchange and silanolic interactions can cause shifts in retention and asymmetrical peaks. The capacity factor and tailing factor of amitriptyline are indicative of the overall performance of the column. Synchronis HPLC columns exhibit excellent peak shape for amitriptyline, demonstrating a highly deactivated silica.

Activity towards chelators

Silica surface metal interactions can cause changes in selectivity and peak shape for analytes which are able to chelate. Changes in the capacity factor and tailing factor of quinizarin, which is a chelator, are indicative of secondary interactions with metallic impurities in the silica. The excellent peak shape observed with Synchronis HPLC columns is indicative of the exceptionally low metals content.

Ion exchange capacity at pH 7.6 (IEC7)

The selectivity factor between benzylamine and phenol is used to measure the total silanol activity on the surface of the silica. At pH>7 the silanol groups are dissociated and combine with the ion exchange sites to influence the retention of benzylamine.



Example chromatogram for Test 2:
Interactions with basic and chelating compounds



Column: Synchronis C18 5 μ m, 100 mm x 4.6 mm
Mobile phase: 10 mM phosphate (pH 7.6):methanol (20:80)
Flow rate: 1.0 mL/min
Temperature: 40 °C
Detection: 254 nm
Injection volume: 5 μ L

1. Theophylline
2. Benzylamine
3. Phenol
4. Quinizarin
5. Amitriptyline

Testing and Quality Control

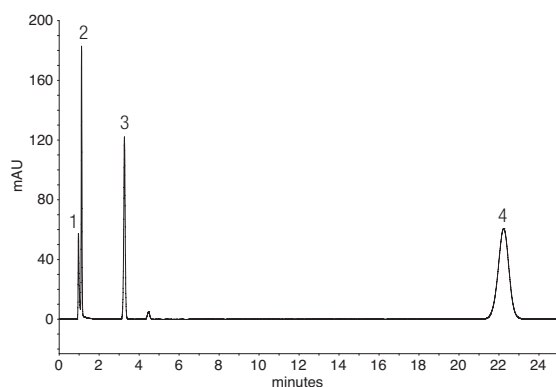
Test 3: Interactions with Acidic Compounds

Activity towards acids

The inertness of the stationary phase towards acidic compounds is also important for consistent retention and selectivity as acidic compounds may become adsorbed onto reversed phase packing materials. The capacity factor and tailing factor of chlorocinnamic acid are measured to test the applicability of the stationary phase to acidic analytes. Synchronis HPLC columns exhibit excellent peak shape, indicating a high degree of inertness towards acidic compounds.

Ion exchange capacity at pH 2.7 (IEC2)

Tanaka¹ showed that the retention of protonated amines at pH<3 could be used to measure the ion exchange sites on the silica surface. Silanol groups (Si-OH) are undissociated at pH<3 and therefore cannot contribute to the retention of protonated amines, but the acidic silanols in the dissociated form (SiO⁻) can. The latter contribute to the retention of the protonated amines. The contribution of the free silanols to retention can be estimated by the selectivity factor between benzylamine and phenol, at pH 2.7.



Column: Synchronis C18 5 μ m, 100 mm x 4.6 mm
Mobile phase: 10 mM phosphate (pH 2.7):methanol (45:55)
Flow rate: 1.0 mL/min
Temperature: 40 °C
Detection: 254 nm
Injection volume: 5 μ L

1. Theophylline
2. Benzylamine
3. Phenol
4. Chlorocinnamic Acid

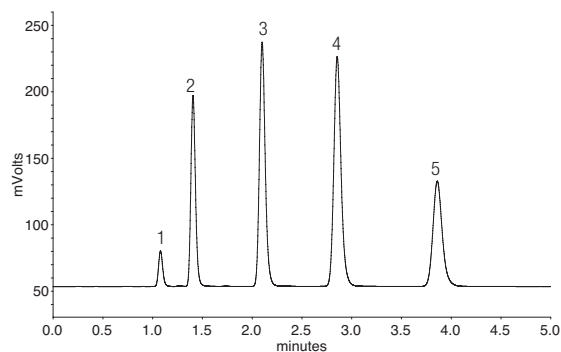
Example chromatogram for Test 3: Interactions with acidic compounds





Test 4: Normal Phase

Unbonded Synchronis silica chromatographic media is a polar stationary phase, which gives excellent reproducibility in normal phase chromatography. Each batch of Synchronis silica is tested under normal phase conditions.



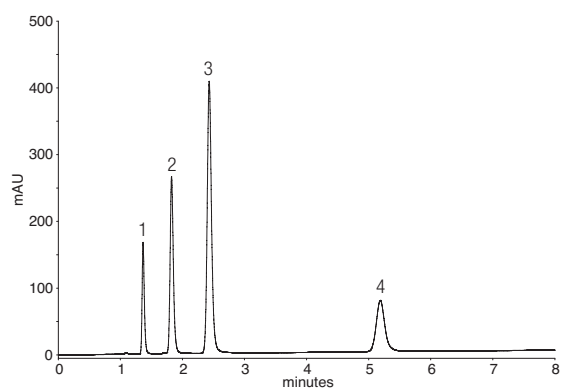
Column: Synchronis Silica 5 μm , 150 mm x 4.0 mm
Mobile phase: iso-octane:ethanol:water (85:14.7:0.3)
Flow rate: 1.25 mL/min
Temperature: 40 $^{\circ}\text{C}$
Detection: 254 nm
Injection volume: 2 μL

1. Toluene
2. Nitrobenzene
3. o-nitroaniline
4. m-nitroaniline
5. p-nitroaniline

Example chromatogram for Test 4: Normal phase retention and selectivity

Test 5: Ion Exchange

Synchronis amino chromatographic media is a polar bonded stationary phase, which gives excellent reproducibility in normal phase chromatography. Each batch of Synchronis amino media is tested under ion exchange chromatographic conditions.



Column: Synchronis Amino 5 μm , 150 mm x 4.6 mm
Mobile phase: 50 mM phosphate (pH 3.0)
Flow rate: 1.0 mL/min
Temperature: 40 $^{\circ}\text{C}$
Detection: 254 nm
Injection volume: 10 μL

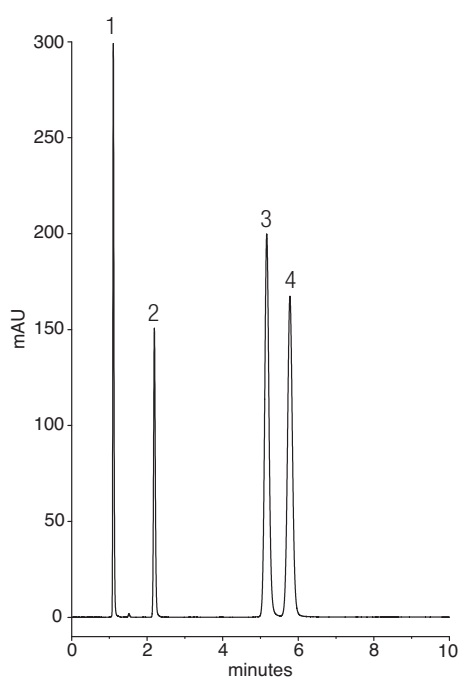
1. Uracil
2. Cytidine monophosphate
3. Adenosine monophosphate
4. Guanosine monophosphate

Example chromatogram for Test 5: Ion exchange retention and selectivity

Testing and Quality Control

Test 6: HILIC Retention and Selectivity

Synchronis HILIC chromatographic media is a polar bonded stationary phase, which gives enhanced retention of polar and hydrophilic analytes. Each batch of Synchronis HILIC media is tested under HILIC conditions.



Column: Synchronis HILIC 5 μ m, 150 mm x 4.6 mm
Mobile phase: 100 mM acetate (pH 5):acetonitrile (10:90)
Flow rate: 1.0 mL/min
Temperature: 40 °C
Detection: 254 nm
Injection volume: 10 μ L

1. Benzophenone
2. Uracil
3. Cytosine
4. Acyclovir

Example chromatogram for Test 6: HILIC retention and selectivity

Column Packing

Every Synchronis HPLC column is individually tested and will not be released unless it meets the required retention, efficiency and peak symmetry. This testing is used to confirm the quality of the column packing process and the stability of the packed bed inside the column.

To ensure the most consistent results column after column, all Synchronis HPLC columns are packed using automated workstations.

To ensure that the measurements are a true indication of the quality of the packing, the testing of individual columns is performed on a highly optimized system.

Summary of Tests Performed on Synchronis HPLC Columns

Test		C18	C8	aQ	Phenyl	Amino	Silica	HILIC
Silica	Particle size and distribution	Y	Y	Y	Y	Y	Y	Y
	Pore size and surface area	Y	Y	Y	Y	Y	Y	Y
Bonded phase	Carbon Load	Y	Y	Y	Y	Y		Y
Chromatographic	Hydrophobic retention	Y	Y	Y	Y			
	Hydrophobic selectivity	Y	Y	Y	Y			
	Steric selectivity	Y	Y	Y	Y			
	Hydrogen bonding capacity	Y	Y	Y	Y			
	Activity towards basic compounds	Y	Y	Y	Y			
	Ion exchange capacity (pH 7.6)	Y	Y	Y	Y			
	Activity towards acidic compounds	Y	Y	Y	Y			
	Ion exchange capacity (pH 2.7)	Y	Y	Y	Y			
	Anion exchange test					Y		
	Normal phase test						Y	
	HILIC retention and selectivity							Y
Column packing	Reversed phase packing test	Y	Y	Y	Y			
	Normal phase packing test					Y	Y	
	HILIC packing test							Y



Synchronis C18

Specifications

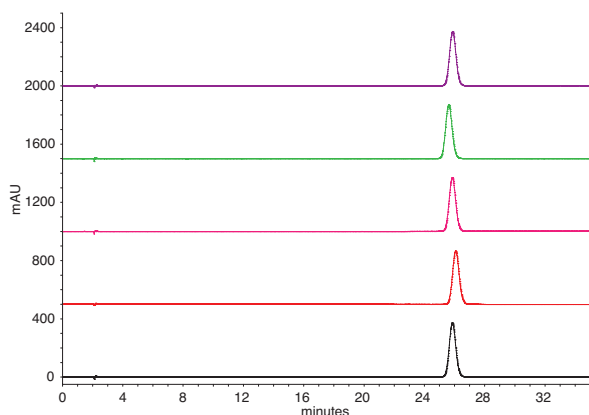
Particle size	1.7 µm, 3 µm, 5 µm	Carbon load	16%
Pore size	100 Å	Endcapped	Yes
Surface area	320 m ² /g	USP classification	L1
pH range	2-9		

Key features/benefits

- Outstanding reproducibility
- Highly pure, high surface area silica
- High carbon load for increased retention
- Double endcapped for extra surface coverage
- Highly inert towards basic compounds
- Rigorously tested to ensure quality

Outstanding column to column reproducibility

When developing a new method, one of the most important goals for the chromatographer is to achieve a consistent, reproducible separation. The selection of a highly reproducible HPLC column is essential if this goal is to be attained.



Synchronis C18 columns show excellent column to column reproducibility

Column: Synchronis C18 5 µm, 250 mm x 4.0 mm
Mobile phase: water:methanol (4:1)
Flow rate: 1.0 mL/min
Temperature: 25 °C
Detection: 265 nm
Injection volume: 10 µL

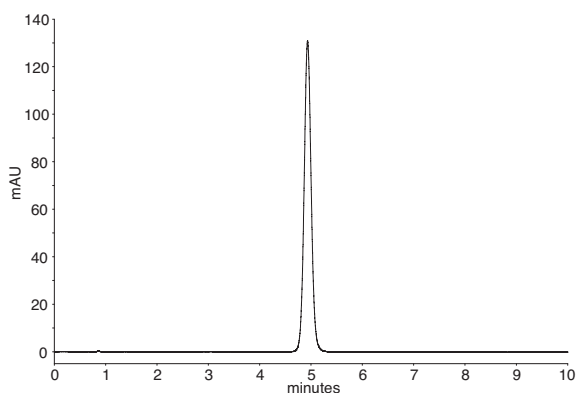
1. Zidovudine

Column	Retention Time (min)	Efficiency	Peak Area
1	25.82	62069	11532105
2	26.03	61688	11543904
3	25.90	62657	11527718
4	25.66	61317	11463444
5	25.92	63142	11520618
Inter-column precision (% RSD)	0.52	1.18	0.27

Synchronis C18 HPLC columns show excellent column to column reproducibility, as illustrated here by the analysis of zidovudine using five separate columns. The reproducibility in terms of retention time and peak area is less than or equal to 0.5 %, column to column. This indicates that the columns are well packed.

The variation in peak area is 0.27 %, which is important for quantitation of analytes.

Application: Chloramphenicol (USP)

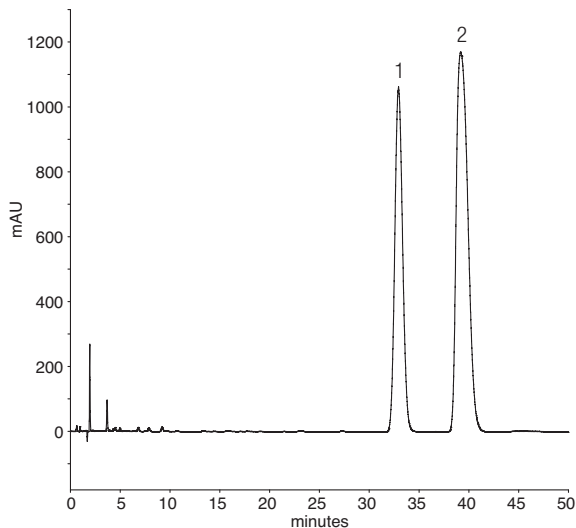


Column: Synchronis C18 5 μ m, 100 mm x 4.6 mm
 Mobile phase: water:methanol:glacial acetic acid (54.9:45:0.1)
 Flow rate: 1.0 mL/min
 Temperature: 25 °C
 Detection: 280 nm
 Injection volume: 10 μ L

1. Chloramphenicol

Parameter	USP specification	Measured (6 replicate injections)
Efficiency (N)	> 1800	6164
Tailing factor	< 2	1.06
%RSD retention time	< 1 %	0.03 %
%RSD peak area	< 1 %	0.32 %

Application: Ibuprofen and Valerophenone (USP)



Column: Synchronis C18 5 μ m, 150 mm x 4.0 mm
 Mobile phase: water/phosphoric acid (pH 2.5):acetonitrile (66.3:33.7)
 Flow rate: 2.0 mL/min
 Temperature: 30 °C
 Detection: 214 nm
 Injection volume: 5 μ L

1. Valerophenone
 2. Ibuprofen

Parameter	USP Specification	Measured Valerophenone (5 replicate injections)	Measured Ibuprofen (5 replicate injections)
Resolution	> 2.0	–	3.56
Relative retention time	~ 0.8	0.84	–
Efficiency (N)	–	8317	5872
Tailing factor	–	1.11	1.38
%RSD retention time	–	0.50 %	0.77 %
%RSD peak area	–	0.50 %	0.27 %

See page 21 for part numbers and ordering information

Synchronis C8

Specifications

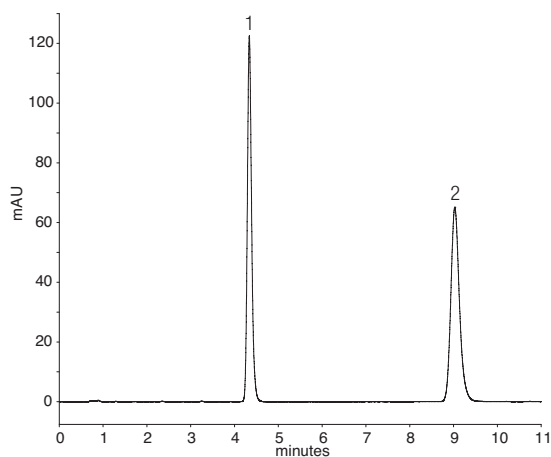
Particle size	1.7 µm, 3 µm, 5 µm	Carbon load	10%
Pore size	100 Å	Endcapped	Yes
Surface area	320 m ² /g	USP classification	L7
pH range	2-8		

Key features/benefits

- Outstanding reproducibility
- Highly pure, high surface area silica
- Less hydrophobic than C18
- Double endcapped for extra surface coverage
- Highly inert towards basic compounds
- Rigorously tested to ensure quality

Synchronis C8 HPLC columns are less hydrophobic than the C18 and are therefore particularly useful where the lesser degree of hydrophobicity is needed in order to successfully retain compounds of interest. Synchronis C8 HPLC columns can also be used where it is desirable to elute compounds more quickly.

Application: Fenoprofen (USP)

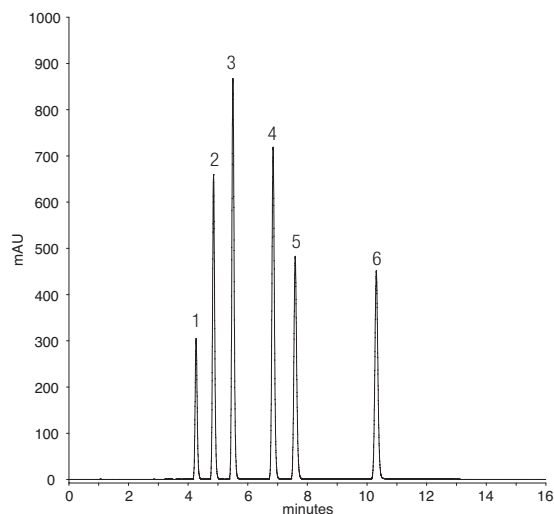


Column: Synchronis C8 5 µm, 150 mm x 4.6 mm
Mobile phase: acetonitrile:water:phosphoric acid (50:49.6:0.4)
Flow rate: 2.0 mL/min
Temperature: 30 °C
Detection: 272 nm
Injection volume: 20 µL

1. Fenoprofen
2. Gemfibrozil

Parameter	USP Specification	Measured Fenoprofen (5 replicate injections)	Measured Gemfibrozil (5 replicate injections)
Resolution	> 8	—	17.6
Relative retention time	~ 0.5	0.48	—
Efficiency (N)	> 3000	9812	10254
Tailing factor	< 2	1.21	1.22
%RSD retention time	< 2 %	0.13 %	0.14 %
%RSD peak area	< 2 %	1.6 %	1.8 %

Application: Uron Herbicides



Column: Synchronis C8 5 μ m, 150 mm x 4.6 mm
Mobile phase: A: water
 B: acetonitrile
Gradient: 35 to 60 % B in 10 minutes
Flow rate: 1.0 mL/min
Temperature: 30 $^{\circ}$ C
Detection: 240 nm
Injection volume: 20 μ L

- 1. Tebuthiuron
- 2. Metoxuron
- 3. Monuron
- 4. Chlorotoluron
- 5. Diuron
- 6. Linuron

Herbicide	RT (%RSD) (6 replicate injections)	Peak Area (%RSD) (6 replicate injections)	Peak Asymmetry
1 - Tebuthiuron	0.31	0.95	1.17
2 - Metoxuron	0.25	0.64	1.18
3 - Monuron	0.18	0.20	1.16
4 - Chlorotoluron	0.12	0.55	1.15
5 - Diuron	0.10	0.37	1.19
6 - Linuron	0.05	0.65	1.13

Synchronis aQ

Specifications

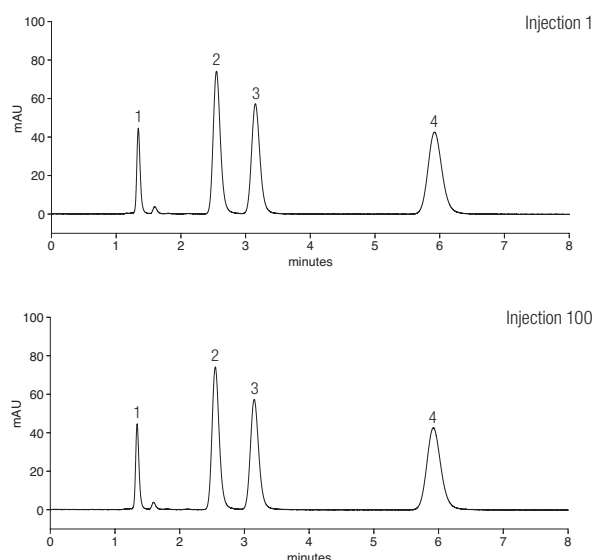
Particle size	1.7 μm , 3 μm , 5 μm	Carbon load	19 %
Pore size	100 \AA	Endcapped	Polar
Surface area	320 m^2/g	USP classification	L1
pH range	2-8		

Key features/benefits

- Stability in 100 % aqueous mobile phase
- Outstanding reproducibility
- Highly pure, high surface area silica
- Highly inert towards basic compounds
- Rigorously tested to ensure quality

The polar endcapping on Synchronis aQ HPLC columns provides a controlled mechanism that retains and resolves polar analytes.

Stability in 100% aqueous mobile phase

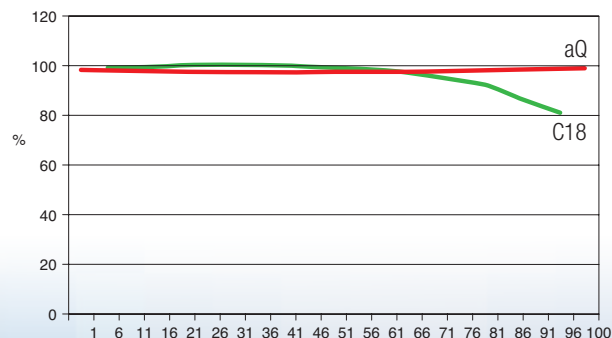


Column: Synchronis aQ 5 μm , 100 mm x 4.6 mm
Mobile phase: 50 mM aqueous potassium hydrogen phosphate (pH 6)
Flow rate: 0.7 mL/min
Temperature: 30 $^{\circ}\text{C}$
Detection: 260 nm
Injection volume: 2 μL

1. Cytidine-5'-diphosphate
2. Adenosine-5'-triphosphate
3. Adenosine-5'-diphosphate
4. Adenosine-5'-monophosphate

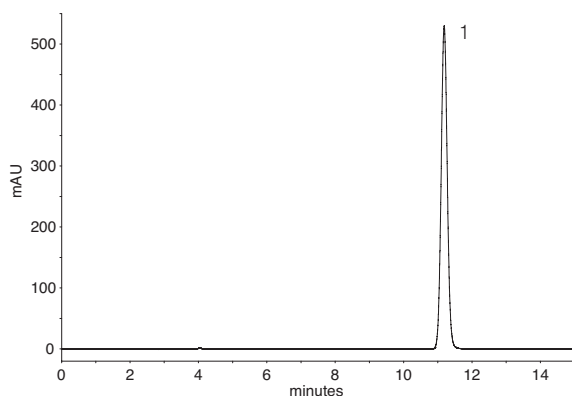
In comparison to a conventionally endcapped C18, the Synchronis aQ polar end-capped C18 stationary phase exhibits superior stability towards aqueous mobile phase. Synchronis aQ shows no degradation in performance after 100 injections in a buffered 100 % aqueous eluent.

In contrast, the performance of the C18 packing begins to deteriorate appreciably after roughly 50-60 replicate injections of the mixture of analytes. The decline in chromatographic performance is more pronounced for the later-eluting compounds. As shown on the right, there is a 20 % decrease in retention time for adenosine monophosphate on the C18 column.



Comparison of relative retention time for 5-AMP on Synchronis aQ and C18 over 100 injections

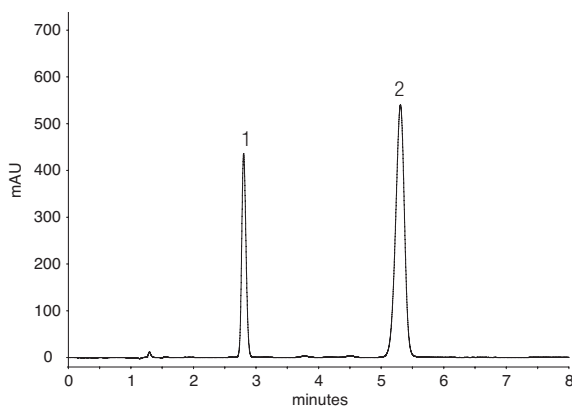
Application: Lamivudine (USP)



Column: Synchronis aQ 5 μm , 250 mm x 4.6 mm
Mobile phase: 25 mM, ammonium acetate (pH = 3.81):methanol (95:5)
Flow rate: 1.0 mL/min
Temperature: 35 $^{\circ}\text{C}$
Detection: 277 nm
Injection volume: 10 μL

1. Lamivudine

Application: Amoxicillin and Potassium Clavulanate (USP)



Column: Synchronis aQ 5 μm , 300 mm x 4.0 mm
Mobile phase: phosphate buffer (pH 4.4):methanol (95:5)
Flow rate: 2.0 mL/min
Temperature: 25 $^{\circ}\text{C}$
Detection: 210 nm
Injection volume: 20 μL

1. Amoxicillin
 2. Potassium Clavulanate

Parameter	USP Specification	Measured Amoxicillin (6 replicate injections)	Measured K Clavulanate (6 replicate injections)
Resolution	> 3.5	–	12.8
Efficiency (N)	> 550	7598	6475
Tailing factor	< 1.5	1.15	0.92
%RSD retention time	< 2 %	0.29 %	0.36 %
%RSD peak area	< 2 %	0.30 %	0.29 %

See page 21 for part numbers and ordering information

Synchronis Phenyl

Specifications

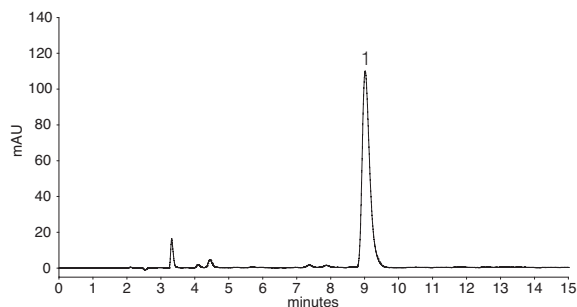
Particle size	1.7 µm, 3 µm, 5 µm	Carbon load	11 %
Pore size	100 Å	Endcapped	Yes
Surface area	320 m ² /g	USP classification	L11
pH range	2-8		

Key features/benefits

- Outstanding reproducibility
- Highly pure, high surface area silica
- Alternative selectivity to C18
- Double endcapped for extra surface coverage
- Highly inert towards basic compounds
- Rigorously tested to ensure quality

Synchronis Phenyl HPLC columns provide alternative selectivity to C18 and are particularly useful for the retention and separation of aromatic compounds.

Application: Oxacillin Sodium (USP)



Column: Synchronis Phenyl 5 µm, 300 mm x 4.0 mm
Mobile phase: phosphate buffer:acetonitrile:methanol (70:30:10)
Flow rate: 1.0 mL/min (2 mL/min in USP method)
Temperature: 25 °C
Detection: 225 nm
Injection volume: 10 µL

1. Oxacillin Sodium (0.11mg/mL)

Parameter	USP Specification	Measured (6 replicate injections)
Efficiency (N)	> 2000	7904
Tailing factor	< 1.6	1.42
%RSD retention time	< 2 %	0.03 %
%RSD peak area	< 2 %	0.29 %



See page 21 for part numbers and ordering information

Synchronis Amino

Specifications

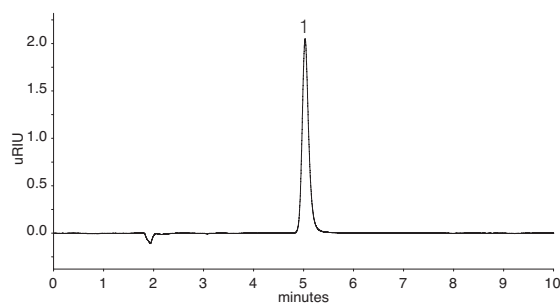
Particle size	1.7 µm, 3 µm, 5 µm	Carbon load	4%
Pore size	100 Å	Endcapped	Yes
Surface area	320 m ² /g	USP classification	L8
pH range	2-8		

Key features/benefits

- Outstanding reproducibility for reversed phase, normal phase, ion exchange and HILIC
- Highly pure, high surface area silica
- Alternative selectivity to C18
- Double endcapped for extra surface coverage
- Rigorously tested to ensure quality

Synchronis Amino HPLC columns exhibit excellent chromatographic properties in weak anion exchange, reversed phase, normal phase and HILIC.

Application: Lactulose



Column:

Mobile phase:
Flow rate:
Temperature:
Detection:
Injection volume:

Synchronis Amino 5 µm, 150 mm x 4.6 mm

water:acetonitrile (30:70)
1.0 mL/min
35 °C
RI
5 µL

1. Lactulose

See page 21 for part numbers and ordering information

Synchronis Silica

Specifications

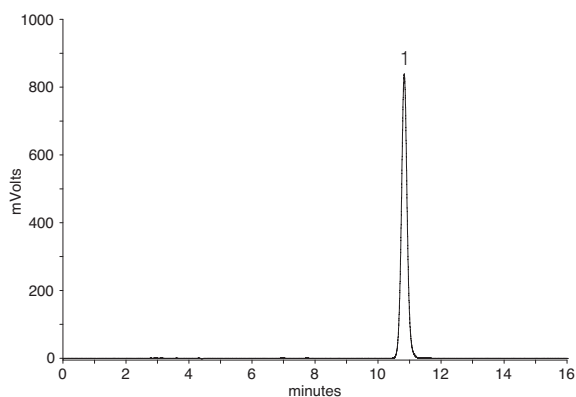
Particle size	1.7 µm, 3 µm, 5 µm	Carbon load	–
Pore size	100 Å	Endcapped	–
Surface area	320 m ² /g	USP classification	L3
pH range	2-8		

Key features/benefits

- Highly pure, high surface area silica
- Excellent reproducibility for normal phase chromatography
- Rigorously tested to ensure quality

Synchronis Silica HPLC columns serve as a powerful and efficient tool for the chromatography of moderately polar organic compounds by normal phase chromatography.

Application: Cetirizine (USP)



Column: Synchronis Silica 5 µm, 250 mm x 4.6 mm
Mobile phase: acetonitrile:water:sulfuric acid (93:6.6:0.4)
Flow rate: 1.0 mL/min
Temperature: 30 °C
Detection: 230 nm
Injection volume: 10 µL

1. Cetirizine

Parameter	USP Specification	Measured (6 replicate injections)
Tailing factor	< 2.0	1.05
%RSD peak area	< 2 %	0.17 %



See page 21 for part numbers and ordering information

Synchronis HILIC

Specifications

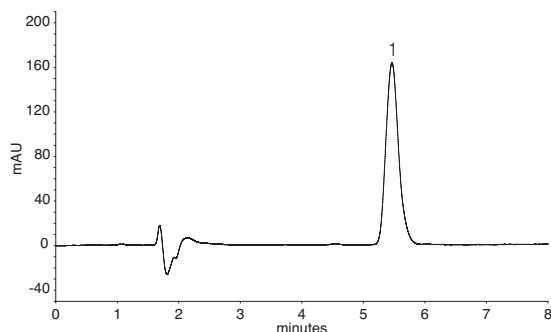
Particle size	1.7 µm, 3 µm, 5 µm	Carbon load	5%
Pore size	100 Å	Endcapped	–
Surface area	320 m ² /g	USP classification	–
pH range	2-8		

Key features/benefits

- Highly pure, high surface area silica
- Zwitterionic bonded phase
- Enhanced retention of polar and hydrophilic analytes
- Excellent reproducibility
- Rapid equilibration
- Rigorously tested to ensure quality

Synchronis HILIC is based on highly pure, high surface area silica particles. The zwitterionic modified stationary phase results in total charge equalisation and therefore a neutral (uncharged) but highly polar surface. Synchronis HILIC HPLC columns offer enhanced retention of polar and hydrophilic analytes.

Application: Allantoin



Column:

Mobile phase:
Flow rate:
Temperature:
Detection:
Injection volume:

Synchronis HILIC 5 µm, 100 mm x 4.6 mm

ammonium formate buffer (pH 3):acetonitrile (10:90)
1.0 mL/min
30 °C
210 nm
10 µL

1. Allantoin

1.7 μm Particles for UHPLC Applications

1.7 μm particles give higher efficiency than 3 μm or 5 μm particles and this efficiency is delivered over a greater range of optimum linear velocity. This makes it possible to operate at higher flow rates without losing performance. Because shorter columns packed with 1.7 μm particles give equivalent efficiency to longer columns packed with 5 μm particles faster analysis and solvent savings for the chromatographer become a reality.

Three Tips for Method Transfer

1. To maintain an equivalent separation when transferring a method it is important to keep the linear velocity constant between the original and new method.
2. Sub-2 μm -based methods are most often transferred to smaller volume columns, so the same injection volume will take up a larger proportion of the new column, possibly leading to band broadening. It is therefore important to scale down the injection volume to match the change in column volume.
3. Geometrical transfer of the gradient requires calculation of the number of column volumes of mobile phase in each segment (time interval) of the gradient in the original method to ensure that the new calculated gradient takes place over the same number of column volumes, for the new column.

System Considerations

With 1.7 μm particles, analyses can be performed with a high linear velocity through the column without loss in performance, provided the LC system is optimized to operate under these conditions. In order to produce fast, efficient chromatography, all system components for the assay should also be considered. Modern ultra high pressure liquid chromatography (UHPLC) instruments, including the Thermo Scientific Dionex™ UltiMate™ 3000 Rapid Separation LC (RSLC) system, will take account of these factors.

There are three major system considerations to remember when using short columns packed with 1.7 μm particles.

1. The system volume (connecting tubing ID and length, injection volume, UV detector flow cell volume) must be minimized.
2. The detector time constant and sampling rate need to be carefully selected.
3. When running fast gradients pump delay volume needs to be minimal.

We also offer a convenient HPLC method transfer calculator at the Chromatography Resource Center
www.thermoscientific.com/crc



Ordering Information

Synchronis HPLC Columns

Particle Size (µm)	Length (mm)	ID (mm)	C18	C8	aQ	Phenyl	Amino	Silica	HILIC
1.7	30	2.1	97102-032130	97202-032130	97302-032130	97902-032130	97702-032130	97002-032130	97502-032130
	50	2.1	97102-052130	97202-052130	97302-052130	97902-052130	97702-052130	97002-052130	97502-052130
	50	3.0	97102-053030	97202-053030	97302-053030	97902-053030	97702-053030	97002-053030	97502-053030
	50	4.6	97102-054630	97202-054630	97302-054630	97902-054630	97702-054630	97002-054630	97502-054630
	100	2.1	97102-102130	97202-102130	97302-102130	97902-102130	97702-102130	97002-102130	97502-102130
	100	3.0	97102-103030	97202-103030	97302-103030	97902-103030	97702-103030	97002-103030	97502-103030
3	30	2.1	97103-032130	–	–	–	–	–	–
	50	2.1	97103-052130	97203-052130	97303-052130	97903-052130	97703-052130	97003-052130	97503-052130
	50	3.0	97103-053030	–	–	–	–	–	–
	50	4.6	97103-054630	–	–	–	–	–	–
	100	2.1	97103-102130	–	–	–	–	–	–
	100	3.0	97103-103030	97203-103030	97303-103030	–	–	–	–
	100	4.6	97103-104630	97203-104630	97303-104630	97903-104630	97703-104630	97003-104630	97503-104630
	150	2.1	97103-152130	–	–	–	–	–	97503-152130
150	4.6	97103-154630	97203-154630	97303-154630	97903-154630	97703-154630	97003-154630	97503-154630	
5	30	2.1	97105-032130	97205-032130	97305-032130	97905-032130	97705-032130	97005-032130	97505-032130
	30	3.0	97105-033030	97205-033030	97305-033030	97905-033030	97705-033030	97005-033030	97505-033030
	30	4.0	97105-034030	97205-034030	97305-034030	97905-034030	97705-034030	97005-034030	97505-034030
	30	4.6	97105-034630	97205-034630	97305-034630	97905-034630	97705-034630	97005-034630	97505-034630
	50	2.1	97105-052130	97205-052130	97305-052130	97905-052130	97705-052130	97005-052130	97505-052130
	50	3.0	97105-053030	97205-053030	97305-053030	97905-053030	97705-053030	97005-053030	97505-053030
	50	4.0	97105-054030	97205-054030	97305-054030	97905-054030	97705-054030	97005-054030	97505-054030
	50	4.6	97105-054630	97205-054630	97305-054630	97905-054630	97705-054630	97005-054630	97505-054630
	100	2.1	97105-102130	97205-102130	97305-102130	97905-102130	97705-102130	97005-102130	97505-102130
	100	3.0	97105-103030	97205-103030	97305-103030	97905-103030	97705-103030	97005-103030	97505-103030
	100	4.0	97105-104030	97205-104030	97305-104030	97905-104030	97705-104030	97005-104030	97505-104030
	100	4.6	97105-104630	97205-104630	97305-104630	97905-104630	97705-104630	97005-104630	97505-104630
	150	2.1	97105-152130	97205-152130	97305-152130	97905-152130	97705-152130	97005-152130	97505-152130
	150	3.0	97105-153030	97205-153030	97305-153030	97905-153030	97705-153030	97005-153030	97505-153030
	150	4.0	97105-154030	97205-154030	97305-154030	97905-154030	97705-154030	97005-154030	97505-154030
	150	4.6	97105-154630	97205-154630	97305-154630	97905-154630	97705-154630	97005-154630	97505-154630
	250	2.1	97105-252130	97205-252130	97305-252130	97905-252130	97705-252130	97005-252130	97505-252130
	250	3.0	97105-253030	97205-253030	97305-253030	97905-253030	97705-253030	97005-253030	97505-253030
250	4.0	97105-254030	97205-254030	97305-254030	97905-254030	97705-254030	97005-254030	97505-254030	
250	4.6	97105-254630	97205-254630	97305-254630	97905-254630	97705-254630	97005-254630	97505-254630	

Other column dimensions are available. Please contact Customer Services for more details

Synchronis HPLC Guard Columns (5 µm Particle Size)

Length (mm)	ID (mm)	Guard Holder	C18	C8	aQ	Phenyl	Amino	Silica	HILIC
10	2.1	852-00	97105-012101	97205-012101	97305-012101	97905-012101	97705-012101	97005-012101	97505-012101
10	3.0	852-00	97105-013001	97205-013001	97305-013001	97905-013001	97705-013001	97005-013001	97505-013001
10	4.0/4.6	850-00	97105-014001	97205-014001	97305-014001	97905-014001	97705-014001	97005-014001	97505-014001

Resources

for Chromatographers

Thermo Scientific Chromatography Columns and Consumables Catalog

This extensive catalog offers 600 pages of proven chromatography tools and product selection guides. Available online, with a robust search tool and optimized for your iPad®.

Visit www.thermoscientific.com/catalog



Chromatography Resource Center

Our web-based resource center provides technical support, applications, technical tips and literature to help move your separations forward.

Visit www.thermoscientific.com/crc



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